

BACTERIAL AETIOLOGY OF OSTEOMYELITIS CASES AT FOUR HOSPITALS OF LAHORE

Faria Malik

Department of Pathology, Fatima Jinnah Medical College, Lahore, Pakistan.

Background: The conclusive diagnosis of osteomyelitis requires isolation of pathogen in aspirate from bone lesion, bone debridement and blood culture. The present research was undertaken to study the microbiological pattern of cases of osteomyelitis reporting to four hospitals in Lahore. **Method:** One hundred and fifty patients of osteomyelitis were selected from outpatient departments and Orthopaedic wards of Lahore General Hospital, Sir Ganga Ram Hospital, Services Hospital and Mayo Hospital, Lahore. Specimens of pus from bone, blood and bone debridement were collected. All samples were inoculated onto two Blood Agar and one MacConkey agar plates. One Blood Agar plate was incubated anaerobically for 48 hours and the other two plates aerobically for 24 hours. Smears were made from samples and stained by the Gram's stain. The colonies obtained were processed according to the technique of Mackie and MacCartney. **Results:** The commonest isolates belonged to the Enterobacteriaceae (32.8%), followed by Staphylococcus aureus (29.5%), Pseudomonas aeruginosa (15.5%), anaerobes (2.6%) and miscellaneous (19.3%). Five (2.7%) anaerobic bacteria were isolated. Anaerobic bacteria were peptostreptococci, peptococci and bacteroides either alone or as a mixed infection. **Conclusion:** The present study highlights the importance of microbiological examination of bone in cases of osteomyelitis. Different types of bacteria either alone or as a mixed infection could be the causative agent(s).

Key Words: Bacteria, Aerobes, Staphylococci, Enterobacteriaceae, Pseudomonas, Anaerobes, Osteomyelitis.

INTRODUCTION

Infections of the bone have been known for a long time. Attempts at understanding the disease began a century ago, when in 1884 Rodet reported to the Academy of Sciences in Paris, his experimental production of haematogenous osteomyelitis in animals by means of intravenous injection of Staphylococci¹. The micrococci were injected into a rabbit which developed typical lesion of osteomyelitis in long bones².

The most common route by which bacteria reach the bone is blood stream^{3,4}. However, traumatic modes as penetrating injury⁵, fractures and intramedullary nailing⁶ and post-surgical complications⁷ have been identified. Intravenous drug users^{8,9} and foreign body presence¹⁰ also predispose to bone infection. The initial diagnosis of osteomyelitis is usually made on physical signs¹¹ and by sonography for early soft tissue changes^{12,13}. Magnetic resonance imaging¹⁴ and bone scans¹⁵ are most sensitive and specific. Conclusive diagnosis requires isolation of pathogen in aspirate from bone lesion, bone debridement and blood culture¹⁵.

In Pakistan, only a few workers have studied the bacteriological pattern of osteomyelitis. The reports from different cities have shown different bacteriological pattern. In

a study at Karachi¹⁶, out of 125 cases, 68.6% were reported to be infected with Staphylococci. Karamat *et al*¹⁷ from Rawalpindi have also reported a high frequency (79%) of Staphylococci, whereas Farooq and Ahmad¹⁸ have reported a very low (37.5%) frequency of Staphylococci. Studies from abroad by Karwowska *et al*¹⁹ Alonge *et al*²⁰ and Lobati *et al*¹⁰ also support the predominant role of Staphylococci in bone infection.

The present research study was undertaken to study the microbiological pattern of cases of osteomyelitis reporting to four hospitals in Lahore.

MATERIAL AND METHODS

The study was conducted at the Department of Microbiology, Postgraduate Medical Institute, Lahore. One hundred and fifty patients were selected from outpatient departments and Orthopaedic wards of Lahore General Hospital, Sir Ganga Ram Hospital, Services Hospital and Mayo Hospital, Lahore.

The patients included were from both sexes and all age groups. The only exclusion criterion was those patients who were on antibiotic therapy.

Specimens of pus from bone, blood and bone debridement were collected. All samples were inoculated onto two Blood Agar and one MacConkey agar plates. One Blood Agar plate was incubated anaerobically for 48 hours and the other two plates aerobically for 24 hours.

Smears were made from samples and stained by the Gram's stain. The colonies obtained were processed according to the technique of Mackie and MacCartney²¹.

RESULTS

The age of the patients ranged from 2–90 years with a mean age (\pm SD) of 28.73 \pm 16.64 years. There were 105 males and 45 females.

There were four specimens of bone curetting, two specimens of blood and 144 specimens of pus. Out of these 150 specimens 186 isolates were identified.

The positivity rate according to various hospitals is shown in Table-1.

Table-1: Distribution of positive cultures according to the hospitals.

Hospital	Tested	Positive	Percentage
Mayo	74	70	93.00
Lahore General	27	26	96.29
Services	28	26	92.85
Sir Ganga Ram	21	18	85.71

The commonest isolates belonged to the Enterobacteriaceae (32.8%), followed by Staphylococcus aureus (29.5%), Pseudomonas aeruginosa (15.5%), anaerobes (2.6%) and miscellaneous (19.3%).

The distribution of isolates according to various age groups is shown in Table-2. Five (2.7%) anaerobic bacteria were isolated. Anaerobic bacteria were peptostreptococci, peptococci and bacteroides either alone or as a mixed infection. The miscellaneous group comprised of Streptococci, Staphylococcus epidermidis, Diptheroids, Micrococci and Bacilli.

DISCUSSION

In the present study, monomicrobial infections in cases of osteomyelitis were seen in two third of patients while in one third cases the aetiology was polymicrobial. Other studies, from Pakistan as well as abroad^{17,22,23}, also report the predominance of monomicrobial aetiology (Table-2). Recent studies^{20,24,25} all report an increasing incidence of polymicrobial infection than the series reported in the past^{26,27} in which mainly monomicrobial infection was common.

Table-2: Comparison showing percentage of monomicrobial/polymicrobial osteomyelitis

Study	Year	Country	Monomicrobial	Polymicrobial
Dendrinos <i>et al</i>	1995	Greece	57.1	42.9
Karamat <i>et al</i>	1995	Pakistan	62.5	12.5
Mousa	1997	Iraq	53.8	46.15
Present study	1999	Pakistan	70.7	29.3

As shown in Table-3, *Staphylococcus aureus* was the predominant isolate (29.56%). Most studies as by Alonge *et al*²⁰, Lobati *et al*¹⁰, Karwowska *et al*¹⁹, Carek *et al*²⁸, Marsh *et al*²⁵ and Karamat *et al*¹⁷ also report *Staphylococcus aureus* as a single organism to be the commonly isolated pathogen from bone infection.

Table-3: Distribution of isolates in various age groups of osteomyelitis patients

Patients			Staphylococcus aureus (%)	Enterobacteriaceae (%)	Pseudomonas aeruginosa (%)	Anaerobes (%)	Miscellaneous (%)
Age group (years)	N o.	No. of Microbes					
<15	8	34	29.41	20.59	17.65	-	32.35
15-29	57	70	32.86	30.0	14.28	2.86	20.0
30-44	36	49	24.49	40.81	14.29	6.12	14.29
45-59	18	21	28.57	38.10	14.28	-	19.05
>60	11	12	33.33	41.67	25.0	-	-
Total	150	186	29.56	32.80	15.60	2.69	19.35

Although *Staphylococcus aureus* remains the most frequent pathogen isolated in bone, the distribution varies from two third *Staphylococcus aureus* to one third Enteriobacteriaceae or one third each of *Staphylococcus aureus*, Enteriobacteriaceae and *Pseudomonas aeruginosa*. This increase in *Pseudomonas aeruginosa* as a significant bone pathogen is related to the increasing nosocomial nature of osteomyelitis²⁹.

Mousa²³ reported a slight predominance in the isolation rate of Enteriobacteriaceae. Even in our study there was a difference of only 3.24% between *Staphylococcus aureus* and Enterobacteriaceae group. Enterobacteriaceae are increasingly common nosocomial pathogens²⁹. The third major group in our study was *Pseudomonas aeruginosa*. *Pseudomonas aeruginosa* remains a severe complication of hospitalization³⁰. The total significant number of cases noted to be infected by the Enterobacteriaceae in the present study could be due to blind therapy for presumptive *Staphylococcal* aetiology, thus eliminating some such cases. Moreover the post-traumatic (69) and post-surgical (48) cases during their hospital stay (114 indoor cases) acquired *Klebsiella* and *Pseudomonas* as additional pathogens in the bone. Mixed infections included upto three isolates belonging to aerobes as *Staphylococci*, Enterobacteriaceae,

Pseudomonas, miscellaneous group or with an anaerobe. Enterbacteriaeiae alone as single organism were isolated in thirty cases and as two organisms in eight cases. Klebsiella was the commonest Entertobacteriaceae to be isolated (21 cases) in single or mixed pattern.

Microorganisms as Staphylococcus epidermidis included in miscellaneous group (5 isolates) have gained importance in periprosthetic infections³¹.

This proves the importance of culturing pus from osteomyelitis cases for aerobes, as well as anaerobes for appropriate management and cure of chronic illness.

CONCLUSION

The present study highlights the importance of microbiological examination of bone in cases of osteomyelitis. Microorganisms could not be detected in only 6.6% cases and one third of cases had a polymicrobial aetiology. Any bacterium, Gram positive or negative, aerobe or anaerobe, either alone or as a mixed infection, could be responsible for osteomyelitis.

The future era with modern high speed travel, warfare, use of implants and prosthetics will add to the load of osteomyelitis.

The clinicians should first obtain a microbiological investigation and then treat their patients to halt the chronic relentless course of this crippling disease.

ACKNOWLEDGEMENT

I am grateful to Dr. N. Rehan, Director Research, PMRC Research Centre, Fatima Jinnah Medical College, Lahore and Liaqat Ali Butt for their support and cooperation.

REFERENCES

1. Nade S. Acute haematogenous osteomyelitis in infancy and childhood. J Bone Joint Surg 1983;65B:109-19.
2. Bick EM. An experimental study on infectious osteomyelitis. In: Bick EM (ed). Classics of Orthopaedics. Philadelphia: JB Lippincott Company;1976:461-2.
3. Glover SC, Padfield C, McKendrick MW, Geddes AM, Dwyer NJP. Acute osteomyelitis in a district general hospital. Lancet 1982;1:609-11.
4. Willis RB, Rozenzwaig R. Pediatric osteomyelitis masquerading as skeletal neoplasia. Orthop Clin North Am 1996;27(3):625-34.
5. Gale W, Scott R. Puncture wound of the foot? Persistent pain? Think of Pseudomonas aeruginosa osteomyelitis. Injury: the Br J Acci Surg 1991;22(5):427-8.
6. Court-brown CM, Keating JF, McQueen MM .Infection after Intramedullary nailing of the tibia. J Bone Joint Surgery 1992;74 B:770-4.
7. Khan G, Hussain A, Rehman M. Infection of the sternum and costal cartilages following median sternotomy: Report of 4 cases. JPMI 1997;11(2):224-9.
8. Boll KL, Jurik AG. Sternal osteomyelitis in drug addicts. J Bone Joint Surg 1990;72B:328-9.
9. Kak V, Chandrasekar PH. Bone and Joint infections in injection drug users. Infect Dis Clin North Am 2002;16(3):681-95.
10. Lobati F, Herndon B, Bamberger D. Osteomyelitis: aetiology, diagnosis, treatment and outcome in a public versus a private institution. Infection 2001;29(6):333-6.

11. Tuson CE, Hoffman EB, Mann MD. Isotope bone scanning for acute osteomyelitis and septic arthritis in children. *J Bone Joint Surg* 1994;76B:306-10.
12. Howard CB, Einhorn M, Dagan R, Nyska M. Ultrasound in diagnosis and management of acute haematogenous osteomyelitis in children. *J Bone Joint Surg [Br]* 1993;75B(1):79-82.
13. Mah ET, LeQuesne GW, Gent RJ, Paterson DC. Ultrasonic features of acute osteomyelitis in children. *J Bone Joint Surg* 1994;76B(6):969-74.
14. Onitsuka H. MRI of bones, joints, and soft tissue. *Asian Med J* 1995;38(9):502-8.
15. Warner WC Jr. Osteomyelitis. In: Crenshaw AH, Daugherty K, Curro C. (eds) *Campbell's Operative Orthopaedics*. 8th Ed. St. Louis: Mosby Year Book; 1992:131-50.
16. Alam SI, Khan KA, Ansari AM, Ahmed A. Etiological study of chronic osteomyelitis in Karachi [Letter]. *J Pak Med Assoc* 1991;41:24
17. Karamat KA, Butt T, Abbas G. Osteomyelitis-prevalence and susceptibility pattern of causative micro-organisms in Rawalpindi/Islamabad area. *Pak J Pathol* 1995;6(2):61-6.
18. Farooq U, Ahmad IF. Bacteriological studies in osteomyelitis at Faisalabad. *J Pak Med Assoc* 1988;38:43-7.
19. Karwowska A, Davies HD, Jadavji T. Epidemiology and outcome of osteomyelitis in the era of sequential intravenous-oral therapy. *Pediatr Infect Dis J* 1998;17(11):1021-6.
20. Alonge TO, Ogunlade SO, Fashina AN. Microbial isolates in chronic osteomyelitis—a guide to management. *Afr J Med Sci* 2002;31(2):167-9.
21. Baird D. Staphylococcus: Cluster-forming Gram-Positive cocci. In: Collee JG, Fraser AG, Tenover BC, Tomasz A (eds). *Mackie and McCartney Practical Medical Microbiology*. 14th Ed. London: Churchill Livingstone; 1996:245-61.
22. Dendrinos GK, Kontos S, Lyritis E. Use of the Ilizarov technique for treatment of non-union of the tibia associated with infection. *J Bone Joint Surg* 1995;77A(6):835-46.
23. Mousa HAL. Evaluation of sinus-track cultures in chronic bone infection. *J Bone Joint Surg* 1997;79B(4):567-9.
24. McNally MA, Small JO, Tofghi HG, Mollan RAB. Two-stage management of chronic osteomyelitis of the long bones: The Belfast technique. *J Bone Joint Surg [Br]* 1993;75B(3):375-80.
25. Marsh DR, Shah S, Elliott J, Kurdy N. The Ilizarov method in nonunion, malunion and infection of fractures. *J Bone Joint Surg [Br]* 1997;79B(2):273-9.
26. West WF, Kelly PJ, Martin WJ. Chronic osteomyelitis: I. Factors affecting the results of treatment in 186 patients. *JAMA* 1970;213(11):1837-42.
27. Kelly PJ, Martin WJ, Coventry MB. Chronic osteomyelitis. II. Treatment with closed irrigation and suction. *JAMA* 1970;213:1843-8.
28. Carek PJ, Dickerson LM, Sack JL. Diagnosis and management of osteomyelitis *Am Fam Physician* 2001;63:2413-20.
29. Gentry LO. Newer concepts in antimicrobial therapy. *Clin Orthop Related Res* 1990;261:23-6.
30. Ostermann PAW, Henry SL, Seligson D. The role of local antibiotic therapy in the management of compound fractures. *Clin Orthop Related Res* 1993;295:102-11.
31. Bukhari SAH, Skinner J, Bentley G. Management of delayed infection after total hip replacement (case report) *Journal of Surgery Pakistan* 2002;7:39-41.

Address for Correspondence:

Dr. Faria Malik, 1/C-III, Hussain Chowk, Gulberg III, Lahore. Tele: +92 42 5762152.

Email: fariamalik188@hotmail.com