HYPERTHERMIA AND RADIATION IN TREATMENT OF HUMAN SQUAMOUS CELL CARCINOMA OF CERVIX XENOGRAFTED IN ATHYMIC MICE

M.W. RANA

Introduction

Cytological screening by the use of the Papanicolaou smear has significantly reduced the number of patients with advanced cervical cancers. Early detection, epidemiology, correct diagnosis and treatment of early stages of cervical lesions have also been effective in control and prevention of cervical cancers. But invasive carcinoma of the cervix is still an important aspect of the cervical cancer problem. The greatest enigma at this time is the etiology, or predisposing factors, i.e., role of male and role of herpes and papilloma viruses in the development of these lesions, which classifies it as a sexually transmissible disease.¹ⁿ⁴ This and other psychological and socio-economic factors might hinder some patients from seeking early detection and prevention. There are still hundreds of cases with advanced lesions reported in literature. Beyond stage II, radiation and chemotherapy are the only treatment and may not be well tolerated by the patient. Serious complications due to radiation therapy occur most frequently in advanced stages of this disease because of high doses of radiation which are required to accomplish tumor control.⁵ It is, therefore, important to seek a broader selection of modes of treatment for this cancer, especially in advanced stages.

It is now well established that hyperthermia kills cells in a manner as predictable and repeatable as radiation and chemotherapy agents.⁶ Unlike ionizing radiation, hyperthermia kills both hypoxic and well oxygenated cells.⁷ Furthermore, neoplastic cells tend to be more sensitive to hyperthermia than do normal cells.⁸ In addition, heat interacts with radiation and chemotherapeutic agents.^{9"10} The role of prostaglandin, prostaglandin analogues, and prostaglandin inhibitors in the treatment of cancer is still controversial. Misonidazol (RO-07--582) has been shown to be an electron affinic hypoxic cell sensitizer.¹¹,¹³ It minimizes the effect of oxygen as a reason for failure in the radiation of hypoxic tumors.

We report here the effect of hyperthermia, radiation and chemotherapeutic agents, used alone or in combination, on human carcinoma of the cervix transplanted into athymic (nude) mice, which may offer a broader selection for the treatment of this lesion in human patients.

From St. Louis University School of Medicine, 1402 South Grand, St. Louis, MO 63104 M.W. RANA, Ph.D. Department of Anatomy and Neurobiology.

Human squamous cell carcinomas of the cervix from patients admitted to St. Louis University Hospitals were used in these studies. Immediately after excision, the tumors were placed in sterile petri dishes. The solid parts of tumor were washed twice in sterile phosphate buffered saline (PBS), then were minced into very small particles, and a small amount of PBS was added. Small quantities (0.1ml) of this tumor mixture were injected through a 15-gauge needle into the left hind leg muscles of 6-8 week old female athymic (nude) mice having Balb/C genetic background. A small piece from each original tumor was processed for histological study. All mice were housed, four per cage, at 80°F in air-tight glass door cabinets protected by laminar air flow, and were given autoclaved chow and water. Most of these mice (70-80%) developed tumors 8-10 weeks after the tumor injections. Histologically, the tumors in mice reached about 1 cm in diameter, the mice were divided into groups for each planned treatment.

In the case of hyperthermia, the hind limbs of the mice having tumors were suspended in a constant temperature hot bath at 43°Cfor 40 minutes. After treatment, the mice were placed under a hot lamp (80°F) to prevent cold exposure. Heat treatments were given twice a week for a total of five treatments.

The radiation treatment was administered with a Siemens Gammatron S teletherapy unit (output 100 rads/min. at 80cm SSD). Auxiliary lead collimating blocks (total height 8 inches) having 1.5cm holes were used to confine the radiation field to the area of the tumor and to minimize the whole body dose. Five fractions of 1500 rads each were delivered over a three week period to the depth of 0.5cm using a single *an fass* port. The mice were treated individually to minimize the chance of a geographic miss. To stabilize, the mice were lightly anaesthetized with 20mg/kg of sodium thiamyll before radiation treatment.

In experiment 1, the mice having tumors were divided into three groups. Group I received heat treatment alone; Group II received heat treatment and radiation; Group III received radiation alone.

In experiment 2, the mice having tumors received 15mg misonidazole subcutaneously and were divided into three groups. Group I received misonidazole alone; Group II received misonidazole and heat treatment, Group III received misonidazole and radiation.

In experiment 3, the mice having tumors were treated with indomethacin (20mg/kg orally) or Ibuprofen (The Upjohn Co., 20mg/kg orally) and were divided into four groups. Group I received indomethacin alone; Group II received indomethacin and heat treatment; Group III received Ibuprofen alone; Group IV received Ibuprofen and heat treatment.

In experiment 4, 16, 16 dimethyl PGE₂ (The Upjohn Co., Kalamazoo, MI), 50ug/kg, was injected intraperitoneally, and the mice were divided into two groups. Group I received the prostaglandin analogue only, and Group II received prostaglandin and hyperthermia.

In experiment 5, the mice having tumors were treated with Adriamycin (0.4mg/kg) and

graded doses of radiation. Group I received 500 rads, Group II received 750 rads, Group III received 1,000 rads, and Group IV received 1,500 rads of radiation in each treatment. This experiment was designed to determine the dose of radiation which could be most effective for these treatments. In our previous studies,⁵ Adriamycin was found to be a very effective radio sensitizer.

Some mice having tumors were not treated and served as controls. In each experiment, mice received bi-weekly treatments for a total of five treatments and were observed for six months after the treatments. The tumors were measured and recorded every week by the same person. (16, 16 diamethyl PGE₂ and Ibuprofen were kindly supplied by Drs. Pike and Lammer of the Upjohn Co., Kalamazoo, MI).

Results

The results of these experiments are shown in tables. In experiment 1 (Table I), the tumors in mice treated with hyperthermia alone regressed continuously and disappeared seven weeks after the treatment. In another experiment where the tumors were somewhat larger (2.0 0.5cm), heat treatment was not so effective. The tumors in these mice continued to grow, although more slowly than those in untreated animals. The tumors in mice treated with hyperthermia and irradiation regressed continually and, except in one mouse, totally disappeared 8 weeks after the treatment. The tumors in this mouse, palpable at this time, disappeared completely 10 weeks after the treatment. The tumors in mice treated with radiation alone also showed regression, except in two mice which did not respond to the treatment. These two mice with large tumors (2.2cm and 1.9cm) died seven weeks after the treatment. Recurrence of tumors was not seen in any of the treated mice until six months after treatment. There was no evidence of radiation sickness in any of the mice.

In experiment 2 (Table II), the tumors in mice treated with misonidazole alone continued to grow. The rate of growth in these tumors was much slower than that in untreated mice. In the group of mice treated with misonidazole and hyperthermia, tumors regressed continually. Eight weeks after the treatment, only one mouse had a palpable tumor. The tumors in mice treated with misonidazole and irradiation showed regression except for one mouse which had a palpable tumor. This tumor also disappeared completely one week later. The effect of misonidazole was more pronounced when combined with radiation than when combined with hyperthermia.

In experiment 3 (Table II), the tumors treated with indomethacin or with Ibuprofen continued to grow. The growth of tumors in mice treated with indomethacin was somewhat slower than in the mice treated wⁱth Ibuprofen. The tumors of mice treated with indomethacin and hyperthermia or Ibuprofen and hyperthermia showed continuous regression. Tumors in all mice disappeared six weeks after treatment except for two mice (one from each group) which died four weeks after treatment. The tumors in these mice also showed relative regression. Some of the mice died during or shortly after heat treatment from dehydration or other causes. These mice were not included in the data presented in results. The mice treated with indomethacin and heat, due to some unknown cause, died six weeks after the treatment.

Because indomethacin and Ibuprofen are inhibitors of prostaglandins, an experiment using prostaglandin analogue 16, 16 dimethyl PGE_2 alone and in combination with hyperthermia was performed. The results of experiment 4 w-ere very inconsistent. The tumors in some mice showed a slight regression during the treatment period, and afterward started to increase in size again. Other mice did not respond to this treatment at all.

In experiment 5 (Table III), the treatment with 1,500 rads was found to be most effective. As the radiation dose was decreased, the relative regression of tumors was also decreased. At 500 rads, no noticeable effect of radiation was found. This experiment was performed the earliest and after this observation, the radiation dose of 1,500 rads was fixed for all the other experiments. As mentioned earlier, this dose did not cause radiation sickness in any of these animals.

Discussion

Besides radiopotentiators and radiosensitizers, beneficial effects of hyperthermia in the treatment of cancer have been shown by many investigators.^{8,14 19} Killing effects of hyperthermia on normal^{8,20} and on neoplastic cells^{21 23} *in vivo* and *in vitro* have been carefully studied. The relatively radioresistant S-phase cells are selectively radiosensitized by heat. Although hyperthermia kills both neoplastic and normal cells, rapidly cycling neoplastic cells tend to be more sensitive to hyperthermia than slowly turning normal cells. Unlike radiation, hyperthermia also kills both hypoxic and well-oxygenated cells.^{7,24} Nutrient deficient cells at low pH are more sensitive to heat.²⁵ These are likely to be hypoxic tumor cells, which may be out of cell cycle. This may constitute a basic rationale for the selective effect of heat on tumor cells.

Several possible mechanisms of hyperthermia cell killing have been proposed. Fajards et al²⁶ reported that hyperthermia produced focal cytoplasmic swelling, rupture of plasma membranes, and peripheral migration of heterochromatin. Westura and Dewey²² indicated that protein molecules are particularly heat sensitive, and damage occurs primarily in chromosomes, at least in the cells engaged in DNA synthesis. Overgaard²⁷ proposed that the damage to the cell is caused by the activation of lysosomes which cause the release of their hydrolytic enzymes into cell cytoplasm. Other observations indicated that heat could increase membrane fluidity and membrane transport, and could decrease all membrane activity.^{9, 28,29}

Whatever the mechanism might be, in our studies heat showed an impressive effect on the tumor size. In one of our experiments where the tumors were large (2cm in diametre) at the beginning of treatment, hyperthermia failed to retard their growth. As the time in treatment was equal for all studies, it may be that the centre of the larger tumors did not reach 43°C during the treatment. Time and temperature relationship is a very important parameter when using heat treatment. Temperature of 43°C has been found to be most effective²⁷,³⁰ During our studies, the centre of the tumor measuring about 1cm in diametre reached 43°C within 15 minutes of treatment. These observations were the basis for our selection of treatment at 43°C temperature for 40 minutes.

Another important factor in heat treatment is thermo-tolerance. The cells can develop resistance to subsequent heat by prior heat treatment.^{31,32} ^Once the cells become thermo-tolerant,

it takes as much as three.pr four days to revert to their normal sensitivity or the decay of thermotolerance. It appeared that the decay of thermo-tolerance occurred faster in cells that were rapidly dividing than those that were not dividing.³³ To avoid thermotolerance, we treated tumors only twice a week. This also provided enough time for the decay of thermo-tolerance if any had developed.

The interaction between heat and irradiation is very complex and has been discussed at length by many investigators.

able-1 EFFECTS OF HYPERTHERMIA, RADIATION AND MISONIDAZOLE ON THE HUMAN CARCINOMA OF CERVIX TRANSPLANTED IN ATHYMIC MICE

		Pre-treatment		Treatment Period		2 Weeks Post Treatment		4 Weeks Post Treatment		6 Weeks Post Treatment		8 Weeks Post Treatment		10 Weeks Post Treatment	
GROUP		Number of mice	Mean size of tumor	Number of mice	Mean size of tumor	Number of mice	Mean size of tumor	Number of mice	Mean size of tumor	Number of mice	Mean size of tumor	Number of mice	Mean size of tumor	Number of mice	Mean size of tumor
Untreated		4	1.3±0.1	4	1.5±0.2	4	1.8±0.3	4	2.0±0.3	4	3.1±1	4	4.1±1.1 ⁸	1	-
Hyperthermia	(A)	4	1.5±0.2	4	1.3±0.2	4	1.0±0.1	4	0.3±0.2	4	0.07 ^t	2	-		-
	(B)	3	2.0±0.5	3	2.0±0.2	3	2.2±0.3	3	2.5±0.3	3	3.0±0.5	3	3.2±0.3		
Radiation		12	1.1±0.3	12	1.1±0.3	11 ^C	1.0±0.3	11	0.9±0.2	11	0.8±0.3	9 ^c	0.4±0.6	9	.04±0.12
Hyperthermia a radiation	and	5	1.2±0.2	5	1.0±0.4	5	0.8±0.6	5	0.4±0.4	4 ^C	0.25±0.1	4	0.1±0.1 ^t	þ	

a - due to large size of tumors mice were sacrificed.

b - only one mouse with tumor

c - mice found dead in the cage.

Table II EFFECT OF HYPERTHERMIA AND THERAPEUTICAL AGENTS ON THE HUMAN SQUAMOUS CELL CARCINOMA OF THE CERVIX TRANSPLANTED IN ATHYMIC MICE

GROUP	Pre-tre	eatment	Treatment Period		2 Weeks P	ost Treatment	4 Weeks P	ost Treatment	6 Weeks	Post Treatment	8 Weeks Post Treatment	
GROOP	No. of Mice	Mean Size of Tumor	No. of Mice	Mean Size of Tumor	No. of Mice	Mean Size of Tumor	No. of Mice	Mean Size of Tumor	No. of Mice	Mean Size of Tumor	No. of Mice	Mean Size of Tumor
Hyperthermia only	4	1.5±0.2	4	1.3+0.2	4	1.0±0.1	4	0.3±0.2	4	0.07a	-	-
Misonidazole alone	6	1.4+0.3	6	1.4±0.2	6	1.6±0.3	6	1.8±0.3	6	2.0±0.4	6	2.5±0.6
Misonidazole and hyperthermia	4	1.4±0.3	4	1.4±0.2	4	1.2+0.2	4	0.8±0.2	4	0.3±0.3	4	0.1a
Misonidazole and radiation	10	1.1 ±0.2	10	0.9±0.2	9Ь	0.3±0.1	9	0.04a		-		-
Indomethacin alone	4	1.0±0.1	4	1.2+0.1	3b	1.4+0.2	3	1.7±0.2	3	1.9±0.3	3	2.5+0.5
Indomethacin and hyperthermia	8	0.9±0.1	8	0.7±0.1	7ь	0.8+0.1	7	0.6±0.03	7C			
Ibuprofen alone	4	1.1 ±0.1	4	1.3+0.1	4	1 4±0.2	4	1.6±0.3	4	2.1 + 0.3	4	2.8±04
lbuprofen and heat	8	1.0+0.1	8	0.8+0.1	8	0.7±0.2	7b	0.6±0.2	7	0.3a		

a - only one mouse with tumor
b - one mouse found dead in cage.
c - all mice were found dead - necroscopy showed no obvious cause.

Table - III EFFECT OF GRADED DOSES OF RADIATION COMBINED WITH ADRIAMYCIN ON HUMAN SQUAMOUS CELL CARCINOMA OF CERVIX TRANSPLANTED IN AHTYMIC MICE

Doses of Radiation	Pre-treatment		Treatment Period		2 Weeks Post Treatment		4 Weeks Post Treatment		6 Weeks Post Treatment		8 Weeks Post Treatment		10 Weeks Post Treatment	
	No. of Mice	Mean size of tumor	No. of Mice	Mean size of tumor	No. of Mice	No. of Mice	No. of Mice	Mean size of tumor	No. of Mice	Mean size of tumor	No. of Mice	Mean size of tumor	No. of Mice	Mean size of tumor
Untreated	5	1.3±0.1	5	1.5+0.3	5	2.0+0.2	5	2.8+0.3	5	3.5±0.8	5	4.3±0.8ª		
500 rads	6	1.3±0.1	5 ^b	1.4±0.2	5	1.2±0.1	5	1 6±0.3	5	2.1+0.7	5	3.4±0.8ª		
750 rads	6	1.5±0.1	5 ^b	1.5±0.1	5	1.5±0.1	5	2.0±0.1	5	2.4±0.2	5	3.5+0.6 ³		
1.000 rads	6	1 3±0.2	6	1.4+0.1	5	1.1+0.1	5 ^C	0.610.2	5	0.3+0.3	5	0.3 ^d		
1,500 rads	6	1.2±0.1	6	1.1 ±0.3	5 ^c	0.5+0.5	5	0.1 ^d ±0.4	5	-		-		

a - due to large size of tumors mice were sacrificed

b — one mouse died due to anesthesia

c — one mouse was found dead in the cage

d — only one mouse with the tumor.

Li et al³⁴ suggested that heat enhances the cell killing effect of radiation even after X-ray treatment. The mechanism of the radio-sensitizing effect of post-irradiation heat treatment may involve the inhibition of potentially lethal damage repair. Hyperthermia inhibits the repair or radiation-induced single-strand breaks and radiation- induced chromosomal aberrations.³⁵⁻³⁷ In other words, heat reduces the repair of sublethal damage and potentially lethal damage produced by radiation. Although Li et al^{34} reported that heat after radiation inhibits the potential lethal damage, and heat treatment before radiation does not, our observations indicated no difference in the sequence. Heat treatment either before (within 30 minutes) or after (within 1 hour) reduced the size of the tumor equally. The maximum toxicity is observed when radiation is delivered simultaneously with heat. i.e., during the heat intervals.³⁸ Our method of combining the two treatments within a short period of time may be very close to the simultaneous treatment. The phenomenon of thermal tolerance during combined heat and radiation treatment is less important for low fractionated radiation doses (200-300 rads) than for high fractionated radiation doses.³⁰ In our studies, the most effective radiation dose was 1,500 rads, which was selected for all the experiments. Overgaard and Nielsen³⁹ observed that with 5 fractions, each consisting of radiation and heat, separated by 24 hours, the thermal enhancement ratio for a TCD50 was no greater than if the same heat dose had been given only once with a single radiation dose. However, when the fractionation interval was increased, supposedly allowing thermal tolerance decay, the effect of subsequent heat dose was greatly increased. Again, the spacing of treatment (twice a week) in our experiments might have avoided the thermal tolerance for heat radiosensitization, because the combination of hyperthermia with irradiation was found to be the most effective in regression of tumors in our investigation.

The cell killing potential of some drugs is enhanced subsequently by hyperthermia.^{40 43} Hyperthermia is advantageous in targeting and localizing the principal effect of the drug, allowing greater tumor cell kill for a given systemic toxicity.

Misonidazole, a 2-nitroimidazole, an electron-affinic hypoxic cell sensitizer was found to be one of the best Flagel compounds for the treatment of tumors.⁴⁴ It is postulated that its toxic effect is due to its reduction product induced by hydrated electrons in anoxia.⁴⁵ Ionized radiation produces a chemical reduction of misonidazole.⁴⁶ It minimizes the effect of oxygen as a reason for failure in the irradiation of hypoxic tumors. The optimal concentration of this drug is reached two hours after its administration.⁴⁷ Thu[^] the radiosensitivity of the tumor is greatly increased when misonidazole is administered to animals before the radiation. In our experiments, the drug was given two hours before the radiation treatment, and the tumors from all the animals, except one tumor which had become very small, disappeared completely. In this study, misonidazole appeared to be an optimum sensitizer. When this drug was used in combination with hyperthermia, the regression of tumors was not as dramatic as when it was used with radiation. Furthermore, the results obtained by heat treatment alone were very similar to these results. It is our suggestion that the effect shown may be due to the heat alone. Obviously for this reason, we did not find any reference in literature where misonidazole was used in combination with hyperthermia.

Anti-inflammatory drugs such as indomethacin at high concentrations may inhibit phosphodiesterases⁴⁸ and allow accumulation of cyclic AMP with inhibition of cell division.^{49,50} High concentrations of indomethacin may reach toxic levels and produce a non-specific inhibition

of cells. Indomethacin given alone significantly slowed the tumor growth rate, but after the drug withdrawal, the tumor growth resumed. This phenomenon was also reported by Kantor and Hampton.⁵¹ Indomethacin at its high concentrations was also not well tolerated by nude mice. Although the size of tumors regressed continuously when used in combination with hyperthermia, hyperthermia may further enhance its toxic effects, because the mice treated with this regimen died four weeks after the treatment.

Another non-steroid anti-inflammatory drug, Ibuprofen, which inhibits prostaglandin synthesis, reduces tumor growth and increases survival time, was used in this study.^{52,54} In addition, the stimulation of cell swelling, cell aggregation, polymorph nuclear leukocyte locomotion, and lysosomal enzyme release in response to chemoattractant were inhibited by Ibuprofen.55 Ibuprofen was found to be less tosic to mice, and when used in combination with hyperthermia gave better results than heat treatment alone or a combination heat and indomethacin. All of the tumors, except in one mouse, regressed four weeks after the treatments. This drug when used alone did not show any significant effects on the growth rate of the tumors.

The precise mechanism of action of prostaglandins is still unclear. Exogenous prostaglandin E was found to inhibit replication of normal, transformed, and malignant cells *in vitro*⁵⁶⁻⁵⁸ Santaro et al⁵⁹ demonstrated that 16, 16-dimethyl PGE₂, a long acting synthetic analogue of PGE₂, inhibited the growth of B-16 adenoma both *in vitro* and *in vivo*. They further demonstrated that systemic administration of dimethyl PGE₂ significantly inhibited tumor growth and prolonged the median survival in these mice. On the other hand, prostaglandins of E series have been shown to inhibit mitogen-induced stimulation, cytolysis, and antibody production by murine and himan lymphocytes,^{60,61} suggesting that prostaglandins will subvert the immunological system. The results in our study were so inconsistent that no logical conclusion could be drawn.

In short, we have shown here some beneficial effects of hyperthermia in the treatment of human squamous cell carcinoma of the cervix transplanted in nude mice, especially when used in combination with radiation or a drug such as Ibuprofen. Although misonidazole produced dramatic results when used in combination with radiation, its results when combined with hyperthermia were not impressive. The results obtained by combination of indomethacin and hyperthermia were also similar to results obtained by using hyperthermia alone. We hope that this study may provide a broader selection for the treatment of this tumor in human patients.

This study was supported by a grant from the American Medical Association. We thank Mr. D. West, Dr. J.E. Pike, and Dr. L.L. Lemmer of the Upjohn Co., Kalamazoo, MI for supplying 16, 16-dimethyl PGE_2 and Ibuprofen. We also thank Mr. Eric Slessinger, Dosimetrist, Radiology Oncology for his technical assistance and Mrs. Janice W. Rana and Ms. Sharon Hughes for the preparation of this manuscript.

REFERENCES

- 1. Christopherson, W.M. and Parker, J.E. A study of the relative frequency of carcinoma of the cervix in the Negro. Cancer 1960; 13:711-713.
- 2. Kessler, II. Human cervical cancer as a venereal disease. Cancer Res. 1976; 36: 783-791.
- 3. Meisels, A. and Morin, C. Human papilloma virus and cancer of the uterine cervix. Gynecol oncol. 1981; 12: 5111-5123.

- 4. Gissman, L., Wolnik, L. and Ikenberg, H. Human papilloma virus type 6 and 11 DNA sequence in genital and laryngeal papillomas and in some cervical cancers. Proc Natl Acad Sci USA 1983; 80: 560-563.
- 5. Xynos, F.R., Benjamin, I., Sapiente, R., Rana, M.W. and Nalesnik, W.J. Adriamycin and hydroxyurea as radiopotentiators in the treatment of squamos cell carcinoma of cervix implanted in nude mice. A preliminary report. Gynecol oncol. 1980; 9: 170-176.
- 6. Dewey, W.C. and Holahan, E.V. Hyperthermia—basic biology. In: Rosenblum ML, Wilson C.B, eds. Progress in Experimental Tumor Research, Brain Tumor Therapy. Switzerland: S. Karger Medical and Scientific Publishers 1984; 28: 198-219.
- 7. Dewey, W.C., Thrall, D.E. and Gillette, E.L. Hyperthermia and radiation. A selective thermal effect on chronically hypoxic tumor cells *in vivo*. Int J Radiol oncol Biol Phys 1977; 2: 99-103.
- 8. Overgaard, K. and Overgaard, J. Investigation on the possibility of a thermic tumor therapy. I. Shortwave treatment of a transplanted isologous mouse mammary carcinoma. Eur J Cancer 1972; 8: 65-78.
- 9. Hahn, G.M., Li, G.C. and Shiu, E. Interaction of amphotericin B at 43°C hyperthermia. Cancer Res. 1977;37:761-764.
- 10. Hall, E.J. and Roizin-Towle, L. Biological effects of heat. Cancer Res. 1984; 44: 4708S-4713S.
- 11. Devenkamp, J. and Stewart, E.A. Sensitization of mouse tumors using fractionated x-irradiation. Brit. J. Cancer. 1978; 37 (Suppl III): 259-263.
- 12. Hill, R.P., Bush, R.S. The effect of misonidazole in combination with radiation dose fractionation. Brit. J. Cancer 1978; 37 (Suppl III): 255-258.
- 13. Sheldon, P.W. and Fowler, J.F. Radiosensitization by misonidazole (RO-07-0582) of fractionated x-ray in a murine tumor. Brit J. Cancer 1978; 37 (Suppl III): 242-245.
- 14. Cavaliere, R., Ciocatto, E.C. and Giovanella, BC. Selective heat sensitivity of cancer cells, biochemical and clinical study. Cancer 1967; 20: 1351-1381.
- 15. Connor, W.C., Gerner, E.W., Miller, R.C. and Boone, M.L.M. Prospect for hyperthermia in human cancer therapy. II. Implication of biological and physical data for the application of hyperthermia to man. Radiol. 1977; 123: 497-503.
- 16. Miller, R.C., Connor, W.C., Heusinkveld, R.S. and Boone, M.L.M. Prospect for hyperthermia in human cancer therapy. I. Hyperthermia effect in man and spontaneous animal tumors. Radiol 1977; 1 23: 489-495.
- 17. Marmor, J.B., Hielerio, F.J. and Hahn, G.M. Tumor eradication and cell survival after localized hyperthermia induced by ultrasound. Cancer Res. 1979; 39: 2166- 2171.
- 18. Hall, E.J. Hyperthermia: An overview. Natl Cancer Inst Monogr 61: 15-16, 1982.
- 19. Dewey W.C., Holahan E.V. Thermobiology—rationale for and problems with utilizing hyperthermia in radiotherapy of cancer. Cancer Bull. 1982; 34: 200-208.
- 20. Kase, K. and Hahn, G.M. Differential heat response of normal and transformed human cells in tissue culture. Nature, 1975; 255 : 228-230.
- 21. Heine, U., Sverak, L., Kondratick, J. and Bonai, R.A. The behavior of Hela-Sa cells under the influence of supernormal temperatures. J. Ultrastruct Res. 1971; 34: 375-396.
- 22. Westura. A. and Dewey, W.E. Variation in sensitivity to heat shock during the cell cycle of Chinese hamster cell *in vitro*. Int J. Radiat Biol. 1971; 19: 467-477.
- 23. Palzer, R.J. and Heidelberger, C. Studies on the quantitative biology of hyperthermia killing of Hela cells. Cancer Res. 1973; 33: 415-421.
- 24. Hahn, G.M. Metabolic aspects of the role of hyperthermia in mammalian cell inactivation and their possible relevance to cancer treatment. Cancer Res. 1974; 34: 3117-3123.
- 25. Freman, M.L., Dewey, W.C. and Hopwood, L.E. Effect of pH on hyperthermia cell survival. J. Natl Cancer Irtst. 1977; 58: 1837-1839.
- 26. Fajardo, L.F., Egbert, B., Marmor, J. and Hahn, G.M. Effect of hyperthermia in malignant tumor,

Cancer 1980; 45: 613-623.

- 27. Overgaard, J. Influence of extracellular pH on the viability and morphology of tumor cells exposed to hyperthermia. J. Natl Cancer Inst. 1976; 56:1243-1250.
- 28. Bowler, K., Duncan, C.J., Gladwell, R.T. and Davison, T.F. Cellular heat injury. Comp Biochem Physiol. 1973; 45: 441-450.
- 29. Li, G.C. and Hahn, G.M. Thermal tolerance and tolerance of adriamycin induced by ethanol. Nature 1978; 274: 699-701.
- 30. Dewey, W.C. Interaction of heat with radiation and chemotherapy. Cancer Res. 1984;44: 4714S-4720S.
- Harris, M. Temperature-resistant variants in clonal population of pig kidney cells. Exp Cell Res. 1967; 46: 301-314.
- 32. Field, S.B. and Law, M.P. The response of skin to fractionated heat and x-rays. Brit J. Radiol, 1978; 51: 221-222.
- 33. Gerweck, L.E. and DeLaney, T.F. Persistance of thermotolerance in slowly proliferating plateau phase cells. Radiat Res. 1984; 97: 365-372.
- 34. Li, G.C., Evans, R.C. and Hahn, G.M. Modification and inhibition of repair of potentially lethal x-ray damage by hyperthermia. Radiat Res. 1976; 67: 491-501.
- 35. Corry, P.M., Robinson, M.S. and Getz, B.S. Hyperthermic effects on DNA repair mechanisms. Radiology, 1977; 123: 475-485.
- 36. Dewey, W.C., Sapareto, S.A. and Betten, A. Hyperthermic radiosensitization of synchronous Chinese hamster cells relationship between lethality and chromosomal aberrations. Radiat Res. 1978; 76: 48-59.
- 37. Jorritsma, J.B.M. and Konings, A.W.T. Inhibition of radiation-induced strand breaks by hyperthermia and its relationship to cell survival after hyperthermia alone. Int J. Radiat Biol. 1983; 43: 505-516.
- Dewey, W.C., Freeman, M.L., Raaphorst, G.P., Clark, E.P., Wong, R.S.L., High-field D.P., Spiro, I.J., Tomasovic, S.P., Denman, D.L. and Coss, R.A. Cell biology of hyperthermia and radiation. In: Meyv RE, Withers HR, eds. Radiation Biology in Cancer Research. New York, Raven Press, 1980; 589-621.
- 39. Overgaard, J. and Nielson, O.S. The importance of thermotolerance for the clinical treatment with hyperthermia. Radiother oncol. 1983; 1: 167-178.
- 40. Hahn, G.M., Braun, J. and Har-Kedar, I. Thermochemotherapy: Synergism between hyperthermia (42-43 C)and adriamycin (or bleomycin) in mammalian cell inactivation. Proc Natl Acad Sci USA. 1975; 72: 937-940.
- 41. Hahn, G.M. and Strande, D.P. Cytotoxic effects of hyperthermia and adriamycin on Chinese hamster cells. J Natl Cancer Inst. 1976; 57: 1063-1067.
- 42. Hahn, G.M. Hyperthermia and Cancer. New York, Plenum Publishing Corp. 1982; 74-86.
- 43. Roizin-Towle, L., Hall, E.J. and Capuano, 1. Interaction of hyperthermia and cytotoxic agents. Natl Cancer Inst Monogr. 1982; 61: 149-151.
- 44. Fowler, J.E., Adams, G.E. and Devenkamp, J. Radiosensitizers of hypoxic cells in solid tumors. Cancer Treatment Rev. 1976; 3: 227-256.
- 45. Wada, J., Suzuki, T. and Iwasaki, M. A new non-steroidal anti-inflammatory agent. 2-substituted 5 or 6-benzothiozoleacetic acids and their derivations. J Med Chem. 1973; 16: 930-934.
- 46. Knight, R.C., Rowley, D.A., Skolimowski, I. and Edwards, D.I. Mechanism of action of nitromidazole antimicrobial and anti-tumor radiosensitizing drugs effects of reduced misonidazole on DNA, Int. J. Radiat Biol. 1979; 36: 367-377.
- 47. Sugahara, T. and Nakatsugawa, S. Radiation sensitization studies in Japan. Cancer Treatment Rev. 1981; 8: 51-61.
- 48. Ciosek, C.P., Ortel, R.W., Thanass, M.M. and Newcombe, D.S. Indomethacin potentiates PGE2

stimulated cyclic AMP accumulation in human synoriocytes. Nature London, 1974; 251: 148-151.

- 49. Hail, V., Horokova, S., Shaff, R.E. and Beaven, M.A. Alteration of tumor growth by aspirin and indomethacin: Studies with two transplantable tumors in mouse. Eur J Pharmacol. 1976; 37: 367-376.
- 50. Karamali, R. Prostaglandins and cancer: A review. Prostaglandins and Medicine, 1980; 5: 11-28.
- 51. Kantor, H.S. and Hampton, M. Indomethacin in submicromolar concentrations inhibits cyclic AMP-dependent protein kinase. Nature, 1978; 276: 841-842.
- 52. Magluilo, E., Genzi-Cipelli, R., Lago, A., Azzini, M., Mogin, E. and Gallico, S. Inhibition effect of non-steroidal anti-inflammatory drugs on the chemotaxis of human leukocytes in vitro. Int J Clin Pharmacol Biopharm, 1977; 15: 417-418.
- 53. Bennett, A., Charlier, E.M., McDonald, A.M., Simpson, J.S., Stamford, K.E. and Zebro, T. Prostaglandins and breast cancer. Lancet, 1977; 2: 624-626.
- 54. Higgs, G.A., Eakins, K.E., Mugridge, S., Moncada, S. and Vane, J.R. The effect of non-steroid anti-inflammatory drugs on leukocyte migrations in carrageenan- induced inflammation. Eur J Pharmacol. 1980; 66: 81-86.
- 55. Maderazo, E.G., Breaux, S.B. and Woronick, C.L. Inhibition of human polymorphonuclear leukocyte cell responses by Ibuprofen. J Phar Sci. 1984; 73: 1403-1406.
- 56. Sykes, J. and Maddox, I. Prostaglandin production by experimental tumors and effect of antiinflammatory compounds. Nature (New Biol). 1972; 237: 59-63.
- 57. Saez, J.M., Evain, D. and Gallet, D. Role of cyclic AMP and protein kinase on the steroidogenic action of ACTH, prostaglandin El and dibutyryl cyclic AMP in normal adrenal cells and adrenal tumor cells from humans. J Cyclic Nucleotide Res. 1978; 4: 311-321.
- 58. Horrobin, D.F. Prostaglandins: Physiology, pharmacology and clinical significance. Montreal, Canada, Eden Press, 1978; 157.
- 59. Santaro, M.G., Philpott, G.W. and Jaffe, B.M. Inhibition of tumor growth *in vivo* and *in vitro* by prostaglandin E. Nature 1976; 263: 777-779.
- 60. Goodwin, J.S. and Webb, D. Regulation of the immune response by prostaglandins. Clinical Immun Immunopath. 1980; 15: 106-122.
- 61. Goodwin, J.S. Prostaglandin E and cancer growth: Potential for immunotherapy with prostaglandin synthetase inhibitors. In: Hersh EM, Chirigos MA, Mastringelo MJ, eds. Augmenting Agents in Cancer Therapy: Progress in Cancer Research and Therapy. 1981; 16: 393-415.

TEST OF GENIUS

Ask a friend to read this sentence slowly:

FINISHED FILES ARE THE RESULT OF YEARS OF SCIENTIFIC STUDY COMBINED WITH THE EXPERIENCE OF YEARS.

Then tell him to count aloud the F's in that sentence. Let him count them only once. How many?

A person of average intelligence finds three F's if you spotted four, you are above average. If you got five, you can turn up your nose at almost anybody. If you caught all six, you are properly a genius, and the question is whether you should spend your time taking tests like this ... v